

# WP3 Biochemical fuel production

Macrofuels plenary meeting  
Wageningen Food and Biobased Research  
Wageningen, The Netherlands  
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## Task 3.1 Fermentation of seaweed syrups to ethanol

Xiaoru Hou, Dimitar B Karakashev, Randi Neerup, Anne-Belinda Bjerre (DTI)

- Aim: To achieve final ethanol concentrations on seaweed syrups higher than 4%

# Seaweed Biomass

- Sugar composition of *Saccharina*



	Temperature (°C)	Glucose (% DM)	Xylose (% DM)	Galactose (% DM)	Arabinose (% DM)	Cellulose (% DM)	Hemicellulose (% DM)
Less-fouling <i>S. latissima</i>	105	5.9	-	0.00	0.00	-	-
	70	6.7	-	0.00	0.00	-	-
	55	7.0	-	0.79	2.21	-	7.63
Heavily-fouling <i>S. latissima</i>	70	2.63	-	0.60	0.98	-	4.48
Unseparated <i>S. latissima</i>	air-dried by SAMS	3.55	-	0.52	1.49	-	3.15



**Sugar content of *Saccharina* is too low**  
**Use sugar-rich Seaweed hydrolysate from previous project**



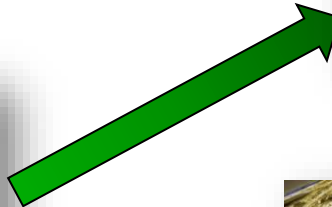
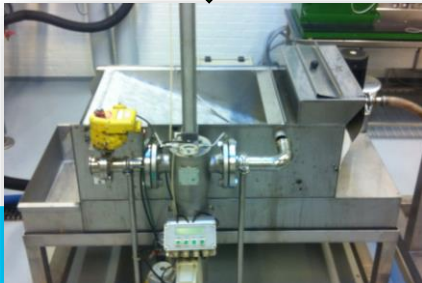
# Seaweed Hydrolysate



**Dried and grinded *L. digitata***  
30 kg  $\approx$  150 kg fresh *L. digitata*



**Enzymatic hydrolysis**  
800L reactor with 600L working volume  
 $T = 45^{\circ}\text{C}$



**Liquid fraction**

Stored at  $T = -20^{\circ}\text{C}$

Glucose (g/L)	Glucan (g/L)	Mannitol (g/L)
9.27	5.76	3.32

This hydrolysate was sent to DLO and to Matis. Used for fermentations at DTI and DLO.

# Task 3.1.1 Mesophile yeast fermentation



**Strain:** *Saccharomyces cerevisiae* (Quick Yeast, Doves Farm Foods Ltd.)

**Medium:** the seaweed hydrolysate

**Temp:** 30°C

**Horizontal rotation rate:** 120 rpm

**Final Ethanol Yield:** 73.3 ( $\pm$  2.1)% theoretical max.

**Final Ethanol Conc.:** 3.47( $\pm$ 0.12) g/L

- Fast and efficient fermentation (almost finished after 24 hours)
- No un-fermented monomer glucose was detected
- No obvious lag phase observed (according to CO<sub>2</sub> production monitored by weight loss)
- Glucan and mannitol were not used

# Task 3.1.2 Thermophile anaerobacter fermentation.

- Hydrolysate toxicity test



- 10 % hydrolysate + 90 % BA medium
- 25 % hydrolysate + 75 % BA medium
- 50 % hydrolysate + 50 % BA medium

### **Preliminary results:**

- Excellent bacterial growth, no inhibition observed
- Waiting for HPLC results of sugars and ethanol

**Basic anaerobic (BA) medium** containing 2 g xylose/L (Angelidaki and Sanders 2004)

**pH:** 6.5-7.0

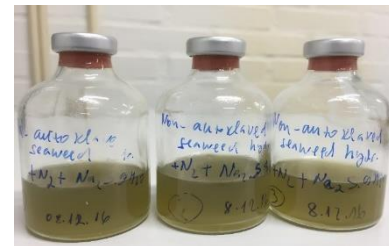
**Temp:** 70°C

**Fermentation hours:** 48-72 h

**Strain:** thermophile *Thermoanaerobacter pentosaceus* DTUO1T, uses both C5, C6 sugars

# Task 3.1.2 Thermophile anaerobacter fermentation.

- Preliminary fermentation on the seaweed hydrolysate
  - ✓ +/- autoclavation
  - ✓ +/- strict anaerobic conditions ( $N_2$  flush versus  $N_2$  flush +  $Na_2S \cdot 9H_2O$ )
- pH 6.5
- Temp: 70°C
- 12 days fermentation (samples collected every 4 days)



Before fermentation



After fermentation

## **Preliminary results:**

-Excellent bacterial growth, no inhibition observed

- Waiting for HPLC results of sugars and ethanol

# Task 3.2. Thermophilic anaerobic biorefinery organisms



Bryndís Björnsdóttir, Guðmundur Óli Hreggviðsson,  
Ólafur H. Friðjónsson, Antoine Moenaert

Engineering of thermophilic strain AK17 for ethanol production and fermentation of laminarin and alginate





# *Thermoanaerobacterium* sp. str. AK17



## Robust thermophilic anaerobe

- $T_{\text{optimal}}$ : 60 °C
- $\text{pH}_{\text{optimal}}$ : 6,0

## Efficient fermentation capacities

- Good ethanol producer
- Acetate and lactate

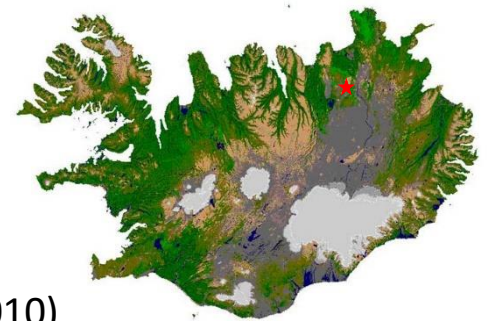


## Diverse metabolic activities and broad substrate range

- Pentose : xylose, arabinose
- Hexose : glucose, galactose, glucuronic acid and mannitol
- Diose : cellobiose

## Susceptible to genetic manipulations

- Natural competence for *Thermoanaerobacterium* genus (Shaw et al, 2010)

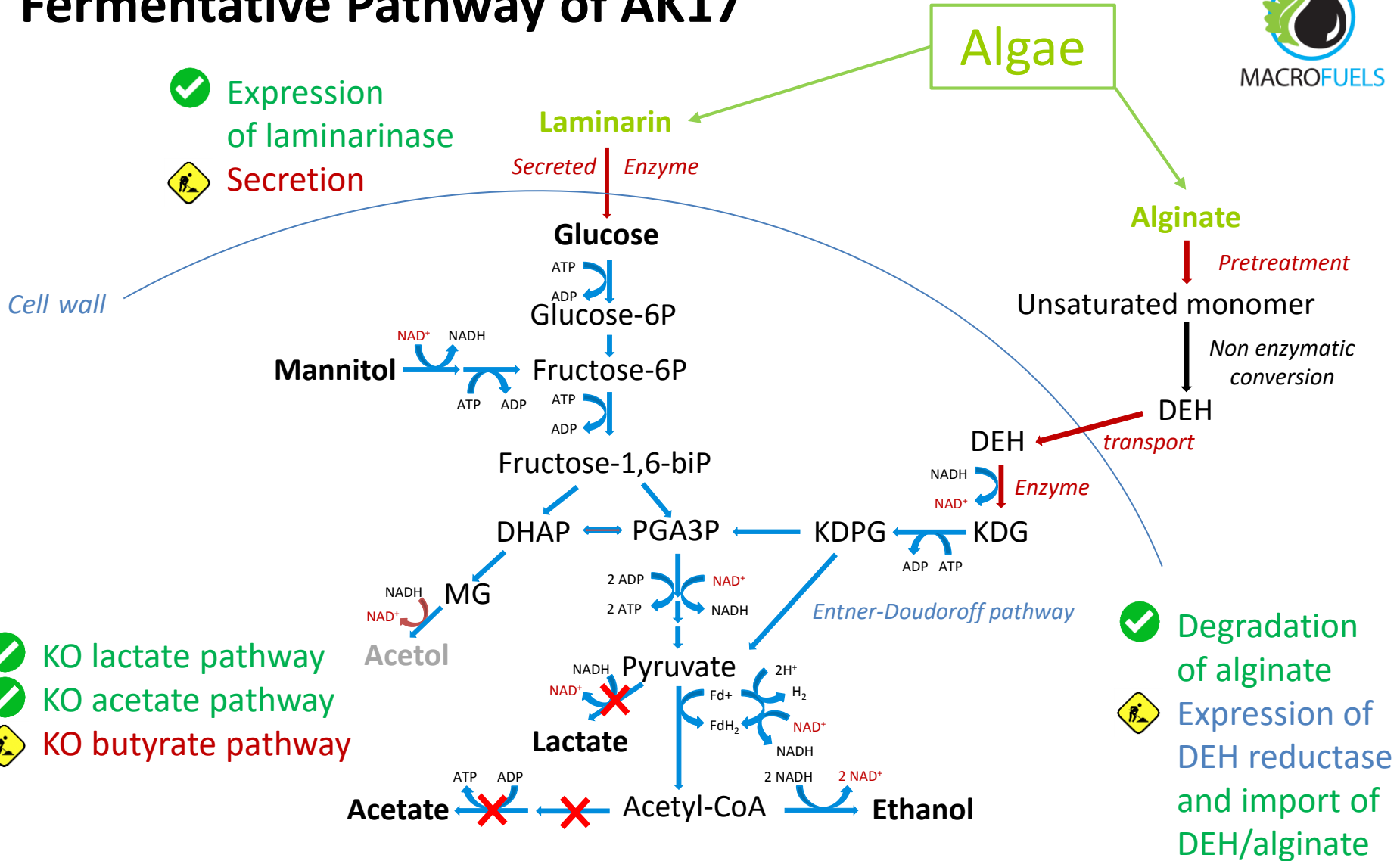


**Aim: Engineer AK17 for macroalgal carbohydrate utilization**



# Fermentative Pathway of AK17

- ✓ Expression of laminarinase
- ⚠ Secretion



- ✓ KO lactate pathway
- ✓ KO acetate pathway
- ⚠ KO butyrate pathway

- ✓ Degradation of alginate
- ⚠ Expression of DEH reductase and import of DEH/alginate

# Task 3.3 Efficient fermentation of seaweed to acetone, butanol and ethanol



Truus de Vrije, Miriam Budde, Hetty van der Wal, Emil Wolbert, Florent Collas, Ana López Contreras

## Seaweed extracts and hydrolysates for fermentation

source of seaweed → preparation of extract/  
hydrolysate

### – Brown seaweed

- *Saccharina latissima* SAMS → WFBR
- *Laminaria digitata* in other project by DTI

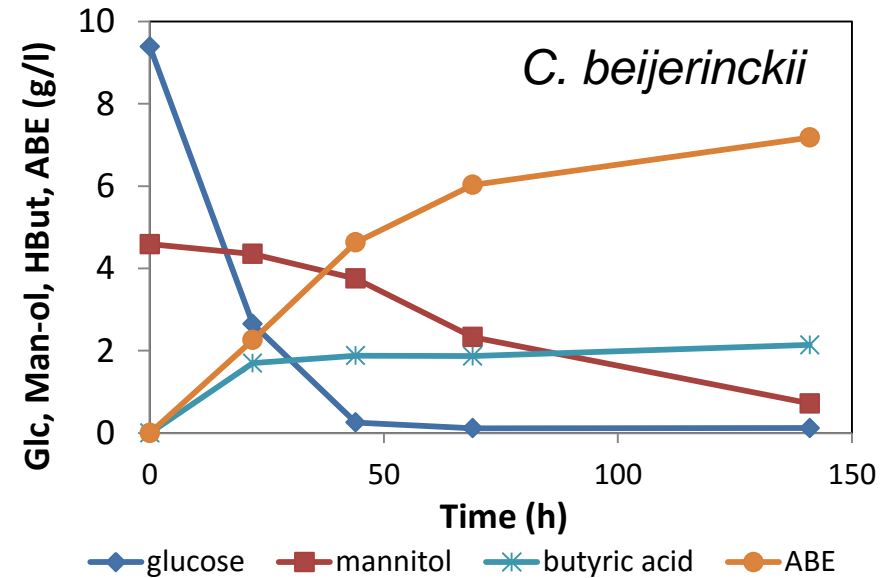
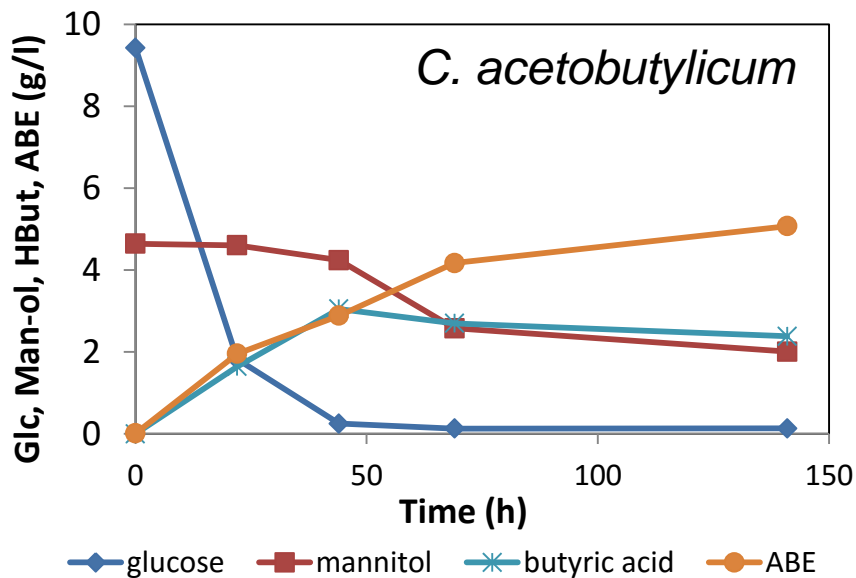
### – Red seaweed

- *Palmaria palmata* SAMS → ECN



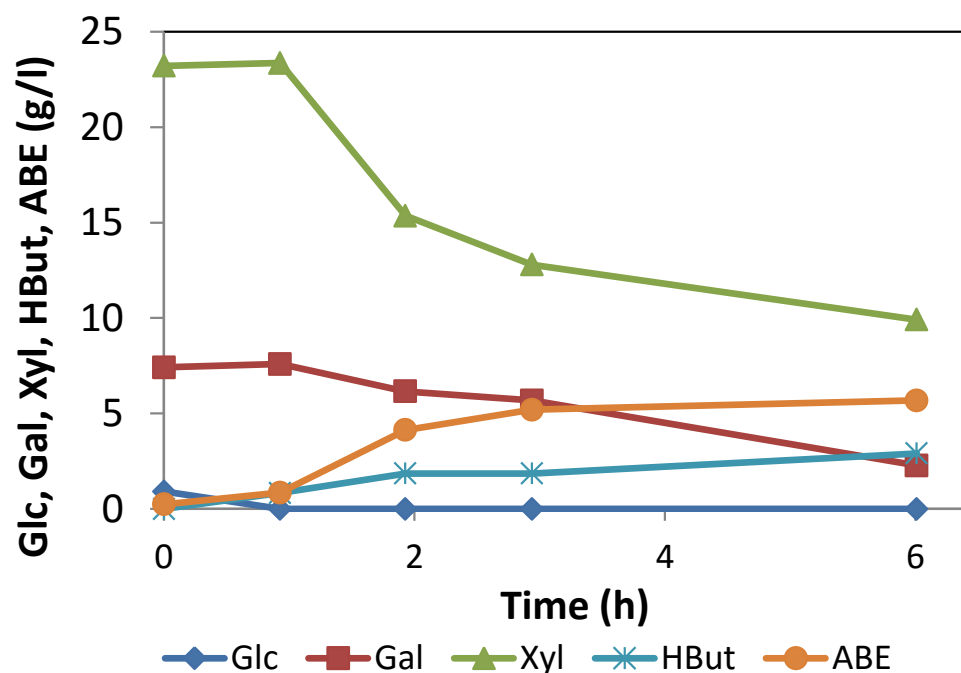
# *Clostridium* spp. with *L. digitata* hydrolysate

- Preferred consumption of glucose over mannitol
- More ABE produced by *C. beijerinckii*
  - growth with HS-medium, without nutrients



# Growth of *C. beijerinckii* on *P. palmata* hydrolysate

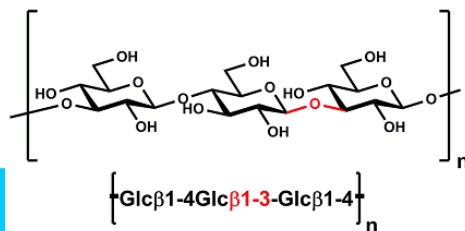
- Preferred consumption of glucose over galactose & xylose



# Direct hydrolysis and fermentation of seaweeds and seaweed fractions

## Approach:

- Tools for genome editing in ABE-producers: CRISPR/Cas9
- Insert enzymes for polymer degradation:
  - Model: CelA (*N. patriciarum*), Family GH6, Binding module CBM1. Active on avicel, CMC and lichenan (Icelandic moss). Under development



# Results & Targets: ABE production



	Aim	<i>Laminaria digitata</i>	<i>Palmaria palmata</i>
Consumed sugars Sugar type	All sugars	Glc, Man-ol Glucan	Glc, Gal, Xyl oligomeric Glc, Gal Xylan
[Sugars], g/l	40 - 60	25	33
Sugar consumption, %	≥ 90	87	60
[ABE], g/l	15 - 20	7.5	5.7
ABE yield, g/g	0.3 – 0.4	0.36	0.28



# Task 3.4 Anaerobic digestion of seaweed SAMS

- Microcosms used to test a range of different conditions
  - Different inoculums (sludge/sediment)
  - Fresh seaweed
  - Pretreatments
  - By products
- Analytical & molecular methods to monitor performance
  - Analytical: methane, total solids, volatile solids, volatile fatty acids
  - Molecular: diversity (gel fingerprint analysis) & quantification (quantitative amplification)
- Scale up best conditions to 10litre
- What do we need to know from you
  - How much sample, when and in what form, composition?



Microcosms

## Analytical



CH<sub>4</sub>, Total & volatile solids, VFA's



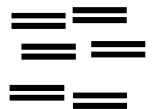
10 litre vessel

## Molecular

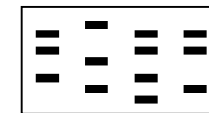
Extract DNA



Amplify DNA & Quantify



Microbial community analysis





# Plans for next 6 months

- **DTI:** Two-stage fermentation of C5-C6-rich seaweed hydrolysate:
  - 1st stage: C6 conversion to ethanol by mesophilic yeast
  - 2nd stage: C5 conversion (effluent from 1st stage after ethanol removal) to ethanol by thermophilic bacterium
- Effect of pH, sugar loading and macronutrients (N, P sources) on thermophilic fermentation

- **MATIS:** Further work on AK17 strain.
  - Fermentation of *Laminaria* hydrolysate
  - Enzyme optimisation
- **DLO:**
  - Fermentation of *Saccharina* hydrolysate
  - Improve conversion of polymers to ABE
- **SAMS:** start preparations for AD of seaweeds and fractions

## Dissemination

- Manuscripts in preparation by Matis, DLO
- Congresses: CBM 2017 (MATIS), Biotech2017 (DLO)

## Next meeting WP3

Skype, in April